

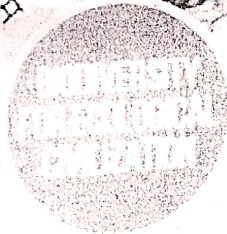


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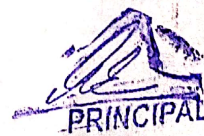
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ISOLATION, IDENTIFICATION AND MORPHOLOGICAL CHARACTERISTICS AND THEIR PHOSPHATE SOLUBILIZING ACTIVITY OF SOIL ALKALIPHILIC ACTINOMYCETES FROM AGRICULTURAL SOIL AT LONAR CRATRE

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ABSTRACT

An alkaliphilic actinomycetes was sequestered from a desert soil sample of Lonar, Dist. Buldhana. The isolate was detected to produce, white, grey, milky white (cotton) colour colonies are obtained from soil sample. These strain produced aerial and substrate mycelium comprising of chain or smooth spore. The colonial growth of strain varied from yellow to grey. All the isolate were later purified and imperiled to a few phosphatic enzymatic screening. Result indicate that number of isolates showed the ability to solubilize phosphate.

INTRODUCTION

Actinomycetes are gram-positive Bacteria viewing a filamentous development like fungi. They are aerobic and extensively present in nature. Actinomycetes are biologically miscellaneous group, as evident by their production of frequent extra cellular enzymes and by the thousands of metabolic yields they produce. Actinomycetes are rich in G+C content with GC% of 57-75%. They are found in dry alkaline soil. Actinomycetes have been well known for the making of secondary metabolites. Many antibiotics are currently used such as streptomycin, tetracycline and erythromycin are the product of actinomycetes. The actinomycetes are important not just in the pharmaceutical industries but also the agriculture. Identification of actinomycetes using microscopic techniques alone was not enough to confirm inevitability. Biological methods would be best method to identify actinomycetes to their type. After isolating an actinomycetes it is primarily acknowledged on the basis of morphological characters so as to have preliminary determination of genus. Microbial natural products have been one of the major incomes for detection of novel drugs. Among the potential sources of natural products, bacteria have been proven to be a prolific source with a surprisingly small group of taxa accounting for the vast majority of compounds discovered. Of the 22,000 recognized bacterial secondary metabolites, 70% are produced by actinomycetes, and two thirds of them are contributed by the genus *Streptomyces* (Subramani and Aalbersberg, 2012). Unlike bacteria, actinomycetes are unique in their morphology with widespread diverging substrate and aerial mycelium bearing chain of arthrospores. The substrate mycelium and spores can be pigmented, which

makes them most colourful and attractive among microbes. On agar plates they form lichenoid, leathery or powdery colonies.

MATERIALS AND METHODS

Method for collection of soil sample

Soil samples were collected about 15 cm below superficial of the soil. All soil samples were collected casually from agriculture Research Center, Lonar. Each sample was occupied from 5-15 cm penetration of the soil by using sterilized degraded metal tube (30 cm length). Soil samples were assorted and sieved to remove stones, leaf, stem and roots. Then, samples were crammed in cleaned and sterile plastic bags, established and stored at 4°C until analysis.


Microbe isolation and Enumeration from soil sample media

Soil samples were air dehydrated for 1 week previous isolation. This helps in decreasing the populace of gram negative bacteria. Soil suspension method described by OSKAY et al (2004) was used, where 19 of the soil. Sample were occupied and mix with 100 ml of sterile distilled water.

The soil suspension was stunned dynamically under room temperature on an orbital shaker at 200 rpm for 1hr. 200 μ l of the soil suspension were pipette and lawn on to Agar.

Isolation of Alkaliphilic Actinomycetes strain from soil sample

Media
The alkaliphilic actinomycetes strains were sequestered using alkaliphilic actinomycetes medium and nutrient agar medium. Alkaliphilic actinomycetes medium was confined, 0.2g sodium casinate, 0.010g L-asparagine, 0.4g sodium propionate, 0.050g dipotassium phosphate, 0.1g


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um sulphate, 0.001g ferrous sulphate, 1.5g agar-
glycerol and PH was adjusted to 9.0

method
method
composed sample, 0.2g of soil samples were
over the surface of media. Then plates were
at 30-35°C for one week. The isolated colonies
checked and respread on designated media.

isolation technique
actinomycetes were isolated from the soil samples by
dilution plate technique (10^{-1} to 10^{-6}). One milliliter
was taken from each dilution and spread evenly
surface of the discriminatory isolation media and
al. Plates encompassing pure cultures were stored
for other examinations. Isolates were used in the
experiments.

Characterization of actinomycetes

Morphological and Cultural Characterization

actinomycetes include abundant order of bacteria,
exhibit wide morphological and physiological
Morphological, physiological and biochemical
of the strains were studied as per International
Project (Shirling and Gottlieb, 1966) and
manual of systematic bacteriology (Williams et
al.).

plates were streaked on to 1) Starch casein agar
0.0 g; casein 1.0 g; K_2HPO_4 0.7 g; KH_2PO_4 0.3
0.4. $7H_2O$ 0.5 g; $FeSO_4 \cdot 7H_2O$) Glycerol
ne agar (ISP 5) (L-asparagine 1.0 g; glycerol 10.0
0.4 g; seawater 1 L; agar 20.0 g; pH 9.2

Culture technique

ip culture is an important tool for learning the
morphology of filamentous actinomycetes under
opted circumstances. Spore chain morphology,
ation of substrate mycelium, aerial mycelium,
d amount of spores in spore chain etc. can be
udied by this method. The isolates were inoculated
rine actinomycete broth and incubated at 28 °C
ys. Plates containing Casein starch peptone yeast
ract agar medium (Casein 3.0 g; maize starch
eptone 1.0 g; yeast extract 1.0 g; malt extract
 $2HPO_4$ 0.5 g; sea water 1 L; pH 7.4; agar 20 g)
anized. Sterile cover slips 3-4 were implanted at
of 45°C into the agar medium. A loopful of spore
ion of actinomycetes was dispensed at the
ion of the medium and cover slip. The plates were
d at 28°C for 4-8 days. The cover slips were
at intervals of 2-4 days and were observed under
ver and oil immersion objectives. Morphology of
mycelium, substrate mycelium, organization of
ous hyphae, their morphology (straight, flexuous,
aped) were recorded according to ISP (Nonomura,
Shirling and Gottlieb,
ected actinomycetes were considered by
ological and biochemical tests [7]. Morphological
mpriprises of macroscopic and microscopic
es The mycelium structure, color arrangements
on the mycelium and colors of colonies were
d and compared with Bergey's manual of
ative bacteriology.

MORPHOLOGICAL CHARACTERISTICS

Morphological characteristics of the strains were
considered according to Methods of Shirling and
Gottlieb, (1966); Bergey's manual of determinative
Bacteriology (Holt et al., 1994) and . The morphology
Of mycelial structures, spore chains and spore surfaces was
observed with a light microscope x 100 (circa 2000, Wolfe
.USA) and Various tests (pigmentation of substrate
mycelium , diffusible pigments were performed for the
characterization of the actinomycetes isolate.

Following are some plates showing growth of
actinomycetes on actinomycetes isolation medium.

The spore chain morphology of actinomycetes developed
in coverslip observed under high power and oil immersion
objectives exposed four types of spore chain morphology.
The most prominent spore chain morphology was the
spiral one, and 34% of the cultures exhibited spiral spore
chain (mostly verticillate type) followed by 28% revealing
rectiflexibiles (straight to flexuous) and 13.9%
retinaculiaperti (open hooks, loops or spirals with one to
two turns) spore chain morphology. (Fig. 2.3 and Fig. 2.4)
Remaining 23.9% of the isolates unveiled long chain of
spores with zigzag fragmenting hyphe

Screening of phosphate solubilizing alkaliphilic actinomycetes

Alkaliphilic actinomycetes were inoculated on starch
casein agar medium encompass 2% of tricalcium
phosphate as a sole phosphorus source for selective
screening of actinomycetes which have capability to
release inorganic phosphate from tricalcium phosphate.
inoculated plates were incubated at 30 degree temp for 7-
10 days. after incubation period plates perceiving zone of
clearance around the colonies which displays the degree
of phosphate solubilization.

RESULT AND DISCUSSION

Isolation of alkaliphilic actinomycetes and distinguished
their morphological and physiological characteristics. A
total of 4 isolates were isolated from the soil samples on
the actinomycetes isolation medium and 2 isolates on the
starch casien agar. Aerial mycelium colours were observed
for the detection of strain varieties in the different soil
samples from dissimilar areas of lonar .colour type were
more plentiful in isolates grown actinomycetes isolation
medium than in isolates grown up on starch casein agar
medium . This medium used for the observation of spores
and sporophores development in actinomycetes it was
noted as medium for characterizing actinomycetes colony
and the colour alteration after the development of
sporulation Regardless the media differences . This data
provided that all the isolates are alkaliphilic organisms.
Alkaliphilic organism is recognized to be able to grow
optimally at PH above 9, usually between 10 & 12
. Nonetheless, it cannot grow or grow slowly at the near
neutral PH value 6.5. zone of clearance around the colonies
which indicates the degree of phosphate
solubilization. Alkaliphilic actinomycetes display
phosphate solubilization activity. The appearance of
colonies ranged from concentric, wrinkled, umbonate, and
chrysanthemum (radial furrows) type (Fig. 2). The spore
mass colour of actinomycetes is considered taxonomic



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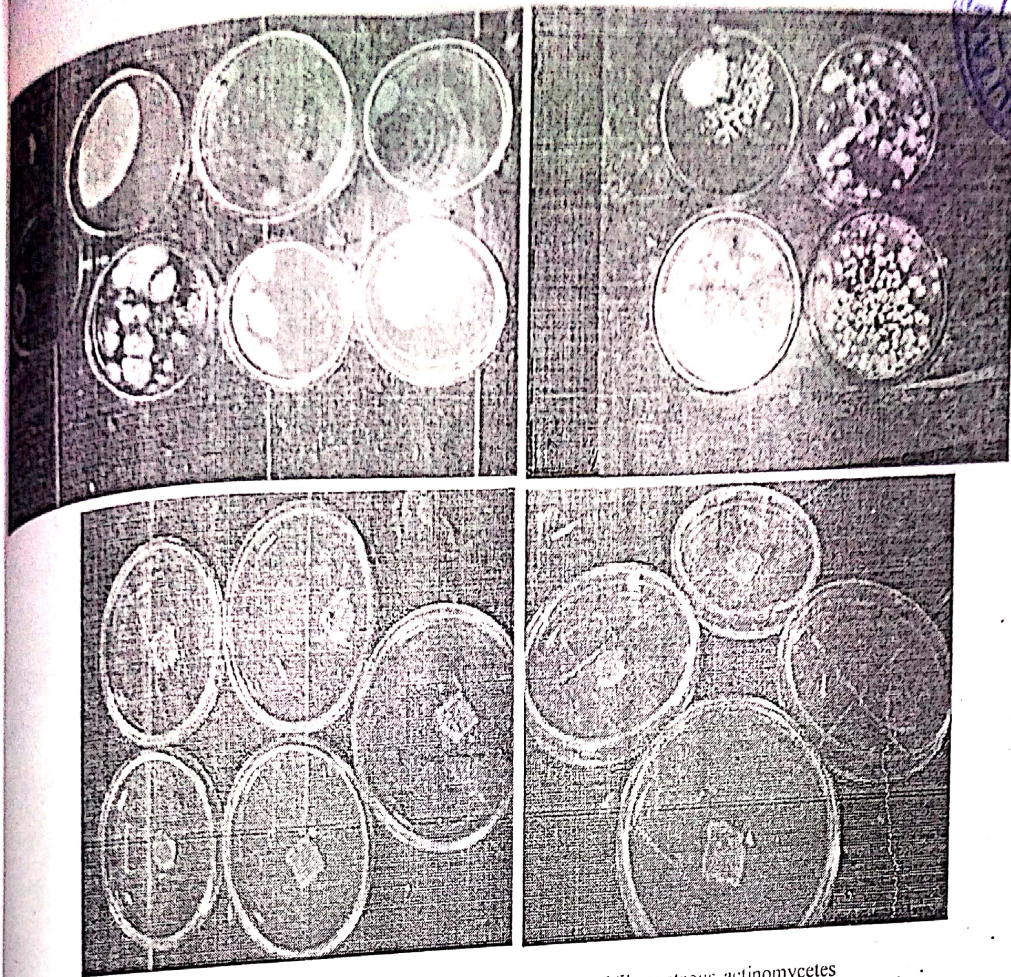


Fig. 1. Cover slip technique for micro morphology of filamentous actinomycetes

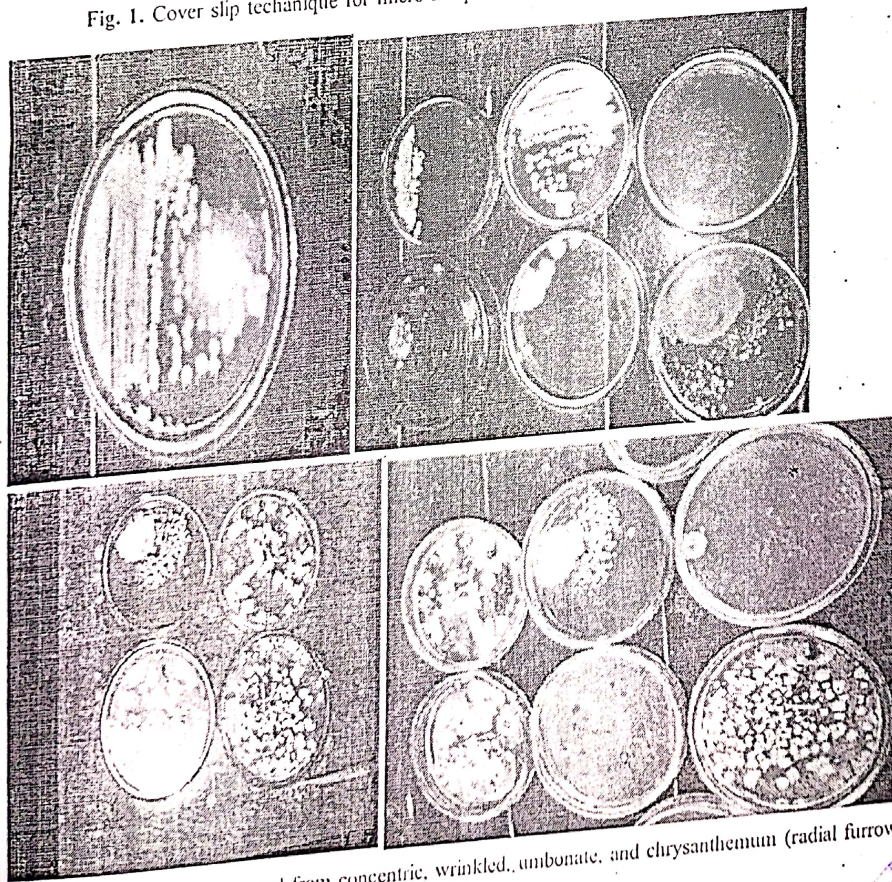


Fig. 2. The appearance of colonies ranged from concentric, wrinkled, umbonate, and chrysanthemum (radial furrows) type.

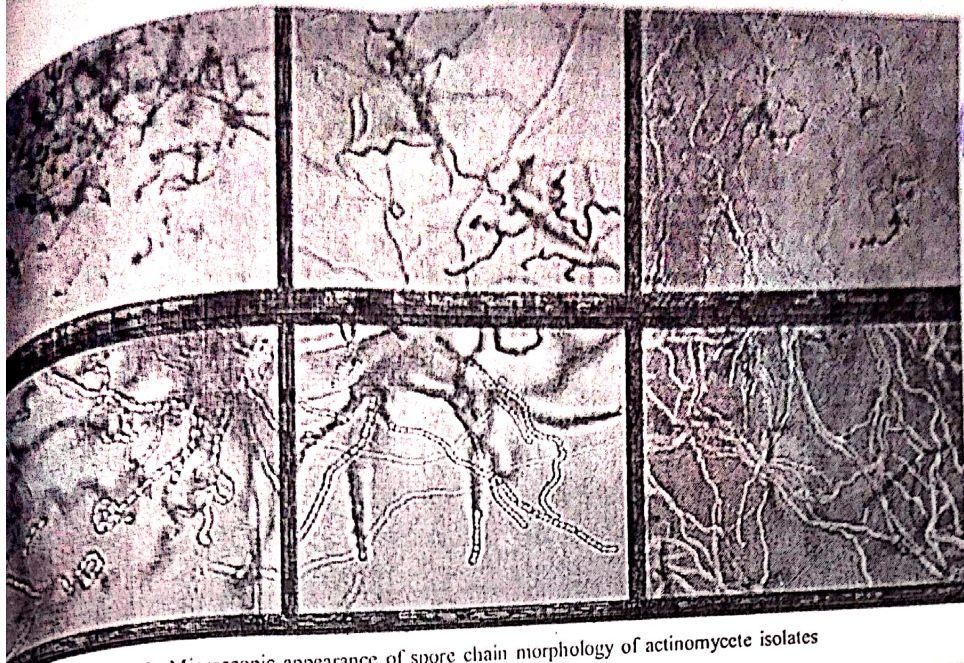


Fig. 3. Microscopic appearance of spore chain morphology of actinomycete isolates

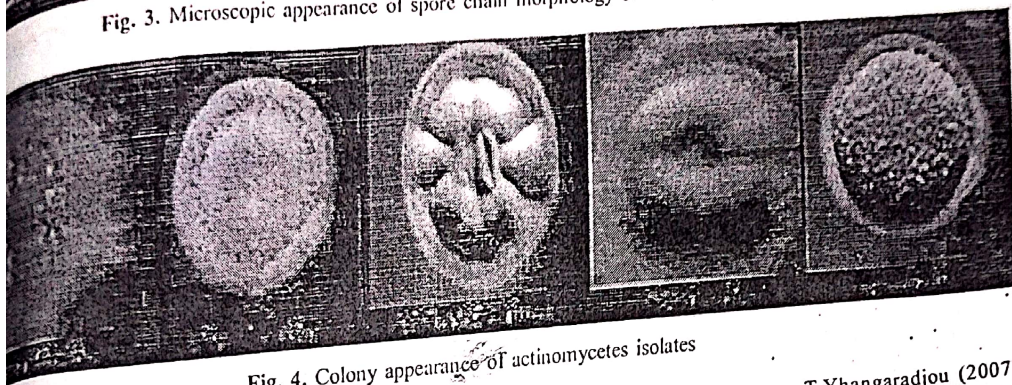


Fig. 4. Colony appearance of actinomycetes isolates

for grouping of actinomycetes in addition, park (1991) reviewed that neutrophilic streptomycetes are able to cultivate between PH 5.0 & 9.0 with growth close to neutrality. The soil samples used in this study were sandy and alkaline (range 7.6 & 8.80). Isolation of actinomycetes from sandy soil has been done with various media.

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